

2. BUCHANAN, T. M., NOMAGUCHI, H., ANDERSON, D. C., YOUNG, R. A., GILLIS, T. P., BRITTON, W. J., IVANJI, J., KOLK, A. H., CLOSS, O., BLOOM, B. R. and MEHRA, V. Characterization of antibody-reactive epitopes on the 65-kilodalton protein of *Mycobacterium leprae*. *Infect. Immun.* **55** (1987) 1000–1003.
3. CONVIT, J., ARANZAZU, N., ULRICH, M., PINARDI, M. E., REYES, O. and ALVARADO, J. Immunotherapy with a mixture of *Mycobacterium leprae* and BCG in different forms of leprosy and Mitsuda-negative contacts. *Int. J. Lepr.* **50** (1982) 415–424.
4. DRAPER, P. Protocol 1/79: Purification of *M. leprae*. Report of the enlarged Steering Committee for Research on the Immunology of Leprosy (IMMLEP) Meeting of 7–8 February 1979. Geneva: World Health Organization, 1979, Annex 1, p. 4.
5. KAPLAN, G. and COHN, Z. A. The immunobiology of leprosy. *Int. Rev. Exp. Pathol.* **28** (1986) 45–78.
6. KLATSER, P. R., DE WITT, M. Y. and KOLK, A. H. J. An ELISA-inhibition test using monoclonal antibody for the serology of leprosy. *Clin. Exp. Immunol.* **62** (1985) 468–473.
7. LAUNOIS, P., NIANG, M. N., DROWART, A., VAN VOOREN, J. P., SARTHOU, J. L., LALU, T., MILLAN, J. and HUYGEN, K. IgG response to purified 65- and 70-kDa mycobacterial heat shock proteins and to antigen 85 in leprosy. *Int. J. Lepr.* **62** (1994) 48–54.
8. MEHRA, V., BLOOM, B. R., BAJARDI, A. C., GRISSO, C. L., SIELING, P. A., ALLAND, D., CONVIT, J., FAN, X.-D., HUNTER, S. W., BRENNAN, P. J., REA, T. H. and MODLIN, R. L. A major T cell antigen of *Mycobacterium leprae* is a 10 kD heat-shock cognate protein. *J. Exp. Med.* **175** (1992) 275–284.
9. RADA, E., SANTAELLA, C., ARANZAZU, N. and CONVIT, J. Preliminary study of cellular immunity to *Mycobacterium leprae* protein in contacts and leprosy patients. *Int. J. Lepr.* **60** (1992) 189–194.
10. RADA, E., ULRICH, M., ARANZAZU, N., SANTAELLA, C., GALLINOTO, M. E., CENTENO, M., RODRIGUEZ, V. and CONVIT, J. A longitudinal study of immunologic reactivity in leprosy patients treated with immunotherapy. *Int. J. Lepr.* **62** (1994) 552–558.
11. RIDLEY, D. S. and JOPLING, W. H. Classification of leprosy according to immunity; a five-group system. *Int. J. Lepr.* **34** (1966) 255–273.
12. ROCHE, P. W., THEUVENET, W. J. and BRITTON, W. Cellular immune responses to mycobacterial heat shock proteins in Nepali leprosy patients and controls. *Int. J. Lepr.* **60** (1991) 36–43.
13. THOLE, J. E. R., JANSON, A. A. M., KIFLE, A., HOWE, R. C., MCLEAN, K., NURILYGN, A., FILLEY, E., SHANNON, E. J., BULLA, G. J., HERMANS, J., DE VRIES, R. R. P., FROMMEL, D. and RINKE DE WIT, T. Analysis of T-cell and B-cell responses to recombinant *M. leprae* antigens in leprosy patients and healthy contacts, significant T-cell responses to antigens in *M. leprae* nonresponders. *Int. J. Lepr.* **63** (1995) 369–380.
14. ULRICH, M., SMITH, P., SAMPSON, C., ZUÑIGA, M., CENTENO, M., GARCÍA, V., MANRIQUE, X., SALGADO, A. and CONVIT, J. IgM antibodies to native phenolic glycolipid-I in contacts of leprosy patients in Venezuela; epidemiological observations and a prospective study of the risk of leprosy. *Int. J. Lepr.* **59** (1991) 405–415.
15. YOUNG, D. B., KAUFMANN, S. H. E., HERMANS, P. W. M. and THOLE, J. E. R. Mycobacterial protein antigens. *Mol. Microbiol.* **6** (1992) 133–145.

Palmo-Plantar Nodular Lesions in Lepromatous Leprosy

TO THE EDITOR:

Mycobacterium leprae has the unique ability among all mycobacteria to utilize DOPA by virtue of the enzyme diphenol oxidase⁽¹⁰⁾. Cells of neural crest origin, e.g., Schwann cells, cells of the adrenal medulla and perineurial cells, can convert tyrosine to DOPA, offering an explanation for unusual tissue specificity of *M. leprae*⁽¹⁰⁾. Another special feature of this organism is that it possesses a distinct predilection for the cooler portions of the body^(2,3,8). Brand⁽³⁾ observed that tendons, cartilage and bones are subject to active infection by *M. leprae* only when they lie close to the

skin where the temperature is low. Based on this concept, the axilla, groin, perineum and narrow transverse band of skin over the lumbosacral region are described as “immune zones” for *M. leprae* because of their relative warmth⁽⁴⁾. Even though palmo-plantar skin is not included in the “immune zones,” its involvement in leprosy is comparatively rare^(1,9,11). All earlier reports on palmo-plantar involvement are in paucibacillary cases^(9,11) and in only a single case of histoid leprosy⁽¹⁾. However, the literature is silent on lepromatous leprosy in this regard. In this communication we are reporting on a lepromatous leprosy patient with nodular lesions over the palms and soles.



FIG. 1. Feet of patient showing plantar nodule lesions.

A 40-year-old male presented to us with "glove-and-stocking" anesthesia and nodular infiltration over the palms, soles and back of elbows (Figs. 1 and 2). The nodules were shiny, smooth, well-demarcated, and 1–2 cm in size, forming almost confluent plaques. The patient also had a few ill-defined partial anesthetic patches on his back and buttocks, infiltration of both earlobes (Fig. 3) and loss of the lateral third of his

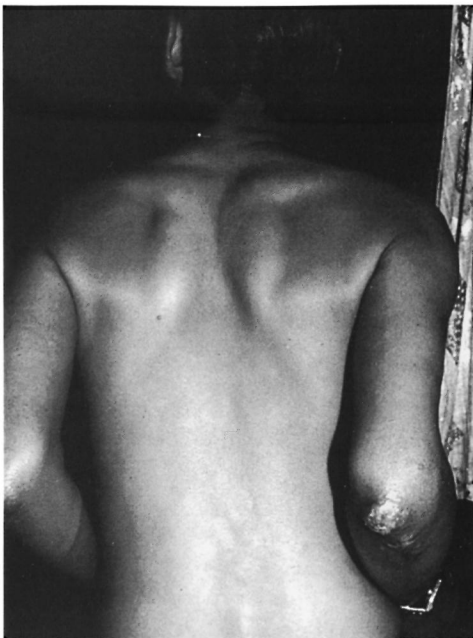


FIG. 2. Back view of patient showing nodules on back of elbow.

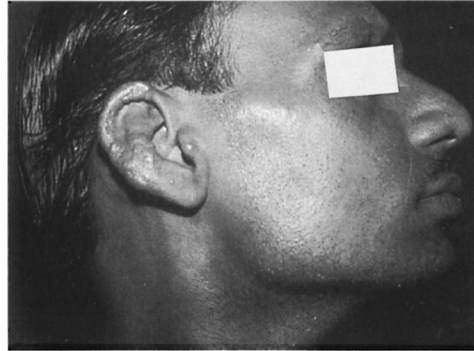


FIG. 3. Side view of patient showing nodular infiltration of earlobe.

eyebrows. The ulnar, common peroneal and greater auricular nerves were enlarged bilaterally and nontender. A bacterial index by slit-skin smear was 5+ from the earlobe, eyebrows and one of the nodules. Histopathology from nodular lesions was consistent with lepromatous leprosy.

Sabin, *et al.* (¹²) have shown that the temperature of the nerve bed is directly related to the depth of the tissues. There is a fairly good amount of fibro-fatty tissue on the palms and soles which ensures an insulating

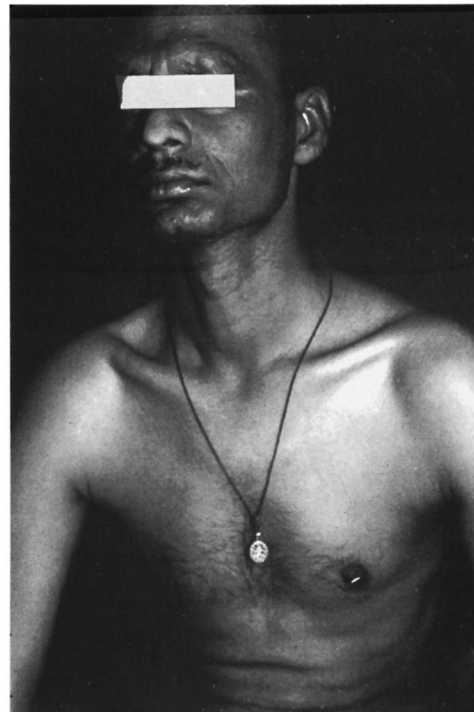


FIG. 4. Frontal view of patient showing chest relatively free from lesions.

property and, hence, a high nerve bed temperature (7). The epidermis is also thicker on the palms and soles compared to other superficial skin areas and, hence, more warm (4). These are probably the reasons for infrequent involvement of the palms and soles in leprosy. However, in our patient the palms and soles were predominantly involved and the classical sites of lepromatous leprosy, including the back, chest, thighs, arms and abdomen, were relatively free. It is possible that the temperature dependency of *M. leprae* is not absolute, as evident by the fact that *M. leprae* survive in warm sites such as bone marrow (13), lymph nodes (6), and liver (5). This reasoning may partly explain the paradoxical distribution of lesions in our patient.

—Uma Shanker Agrawal
Puneet Bhargava
Ram Gulati
Narender Kumar Mathur

Department of Dermatology
SMS Medical College
Jaipur, India

Reprint requests to Dr. N. K. Mathur,
C-24 Peeyush Path, Bapunagar, Jaipur 302
105, India.

REFERENCES

1. BASLAS, R. G., GUPTA, M., ARORA, S. K., MUKHIJA, R. D. and MISRA, R. K. Palmar involvement in histoid leprosy. *Indian J. Lepr.* **64** (1992) 193–195.
2. BINFORD, C. H. Comprehensive program for inoculation of human leprosy into laboratory animals. *Public Health Rep.* **71** (1956) 995–996.
3. BRAND, P. W. Temperature variation and leprosy deformity. In: *VII Transactions of the International Congress of Leprosy, Tokyo, 1958*. Tokyo: Japanese Leprosy Foundation, 1959, pp. 125–129.
4. COCHRANE, R. G. Signs and symptoms. In: *Leprosy in Theory and Practice*. 2nd edn. Cochrane, R. G. and Davey, T. F., eds. Bristol: John Wright & Sons, 1964, p. 266.
5. CONTRERAS, J., JR., TERENCIO DE LAS AGUAS, J. and CONTRERAS, F. Hepatic lesions in lepromatous patients. *Int. J. Lepr.* **37** (1969) 270–279.
6. DESIKAN, K. V. and JOB, C. K. A review of post-mortem findings in 37 cases of leprosy. *Int. J. Lepr.* **36** (1968) 32–44.
7. ENNA, C. D., BERGHOLDT, H. T. and STOCKWELL, F. A study of surface and deep temperatures along the source of the ulnar nerve in the pisohamate tunnel. *Int. J. Lepr.* **42** (1974) 43–47.
8. FELDMAN, W. H. Inducing pathogenesis of *M. leprae* in animals. *Public Health Rep.* **71** (1956) 995.
9. MUKHERJEE, N., GHOSH, S. and KUNDU, S. Palmar lesion in a case of leprosy of the tuberculoid type. (Abstract) *Int. J. Lepr.* **29** (1958) 245.
10. PRABHAKARAN, K. Unique metabolic properties of *Mycobacterium leprae*. *Indian J. Lepr.* **63** (1991) 410–417.
11. RAJENDRA, N. Palmo-plantar lesions in paucibacillary leprosy—unusual clinical expressions. *Indian J. Lepr.* **59** (1987) 188–190.
12. SABIN, T. D., HACKETT, E. R. and BRAND, P. W. Temperatures along the course of certain nerves often affected in lepromatous leprosy. *Int. J. Lepr.* **42** (1974) 38–42.
13. SUSTER, S., CABELLO-INCHAUSTI, B. and ROBINSON, M. J. Nongranulomatous involvement of bone marrow in lepromatous leprosy. *Am. J. Clin. Pathol.* **92** (1990) 797–801.

Posterior Chamber Intraocular Lens Implantation in Smear-Positive Leprosy Patients; a Preliminary Report

TO THE EDITOR:

Cataracts in leprosy patients can a) evolve as a consequence of uveitis, b) manifest as a result of steroid therapy, or c) occur as part of the aging process. Age-related cataract may soon become a leading cause of blindness among leprosy patients in endemic areas.

The best way to rehabilitate patients with age-related cataract is by extracting the cataract and implanting an intraocular lens. There are no reports available in the literature that describe this procedure having

been done on multibacillary leprosy patients whose skin smears were positive for acid-fast bacilli (AFB). We report here three posterior chamber intraocular lens implantations into the eyes of two lepromatous patients whose skin smears demonstrated AFB at the time of surgery.

CASE REPORT 1

A 35-year-old man with lepromatous leprosy, confirmed by histopathology, had completed 2 years of multidrug therapy (MDT). During this time and after the completion of his MDT, he had had several